

The Role of Bacteria in Wetland Functionality and Sustainability

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Important issues in GSL Ecosystem

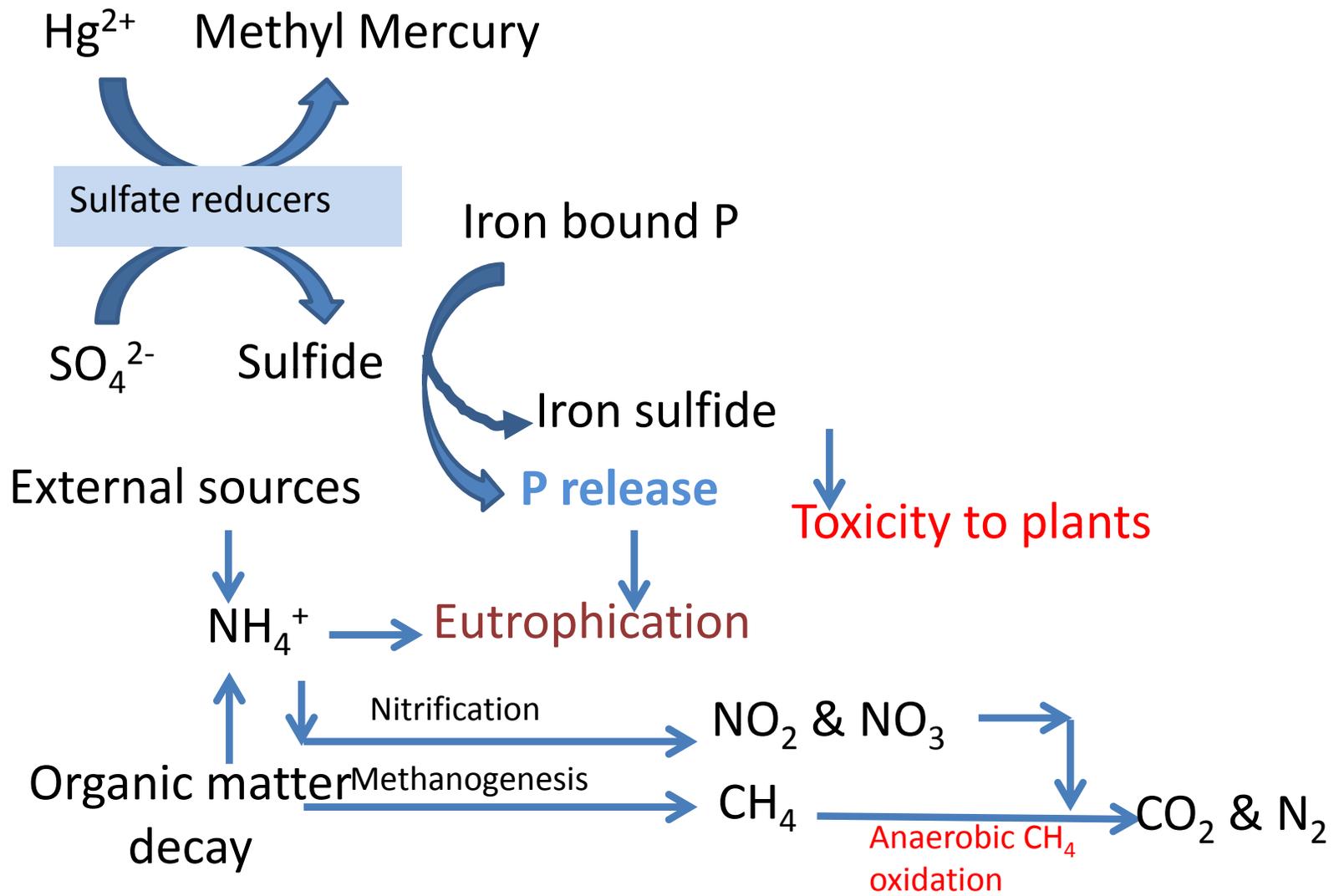
Nutrients

Mercury and Selenium

Salinity gradient

Cyanotoxins

Other heavy metals (??)



Microbial transformations of nutrients and iron dictate vegetation composition in wetlands

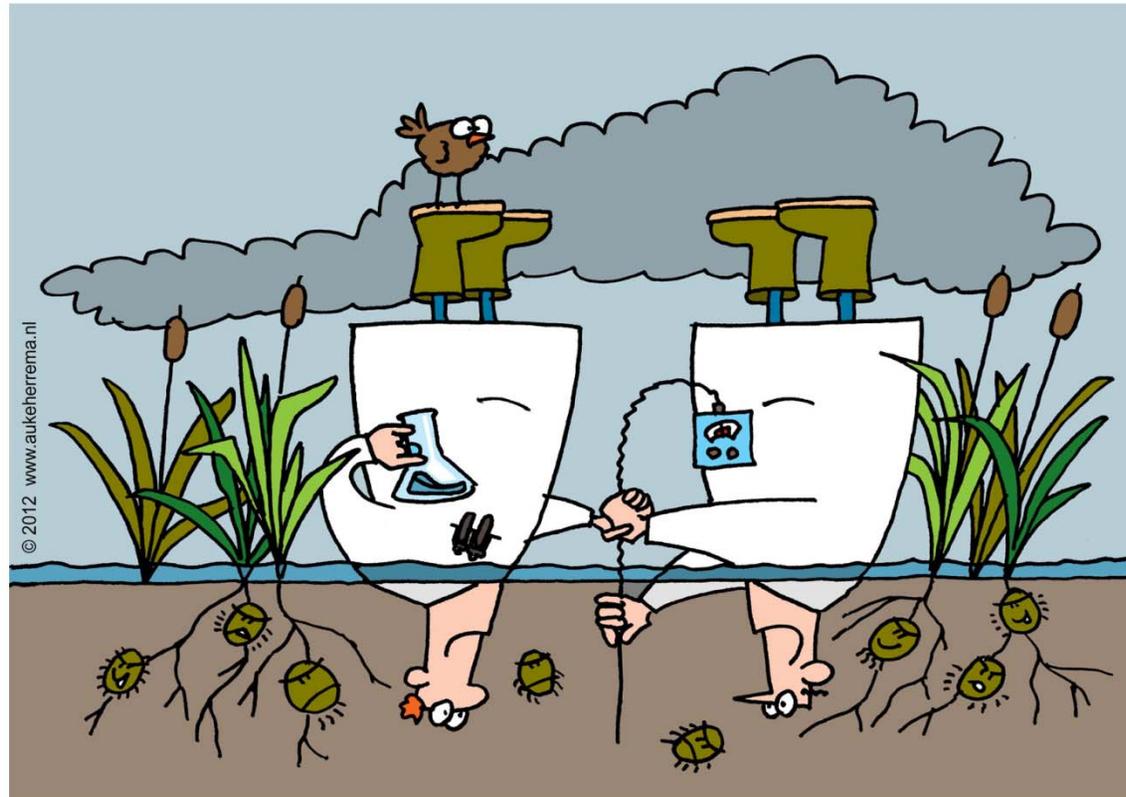


FIGURE showing Below-ground scientific collaboration in wetlands: microbiology meets plant ecology. (Taken from Lamers et al., 2012)

Specific Projects

- Cyanotoxins in Farmington Bay- Central Davis Sewer District
- Mercury and its methylation in GSL- UDWQ
- Nutrient fate flux and fate in Willard Spur Wetlands- UDWQ
- Nutrient fate and dynamics in Farmington Bay Wetlands- USEPA

Should Cyanotoxins be of concern to us?



Algal bloom in Farmington Bay (2008)

- Algal blooms (eutrophication?)
- Cyanobacteria (Blue-green algae) can release Cyanotoxins

**But not all the algal blooms
Correspond to Cyanobacteria**

What are Cyanotoxins?

Cyanotoxins

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graph TD; A[Cyanotoxins] --> B[Hepetotoxins  
(affect liver)]; A --> C[Neurotoxins  
(affect nerve system)]; B --> D[Strains of  
Microcystis, Nodularia, etc.]; C --> E[Strains of  
Aphanizonmenon, Oscillatoria, etc.]; D --> F[Microcystins, Nodularin, etc.]; E --> G[Anatoxins, Saxitoxin, etc.]
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Hepetotoxins
(affect liver)

Neurotoxins

(affect nerve system)



Strains of
Microcystis, Nodularia, etc.

Strains of
Aphanizonmenon, Oscillatoria, etc.



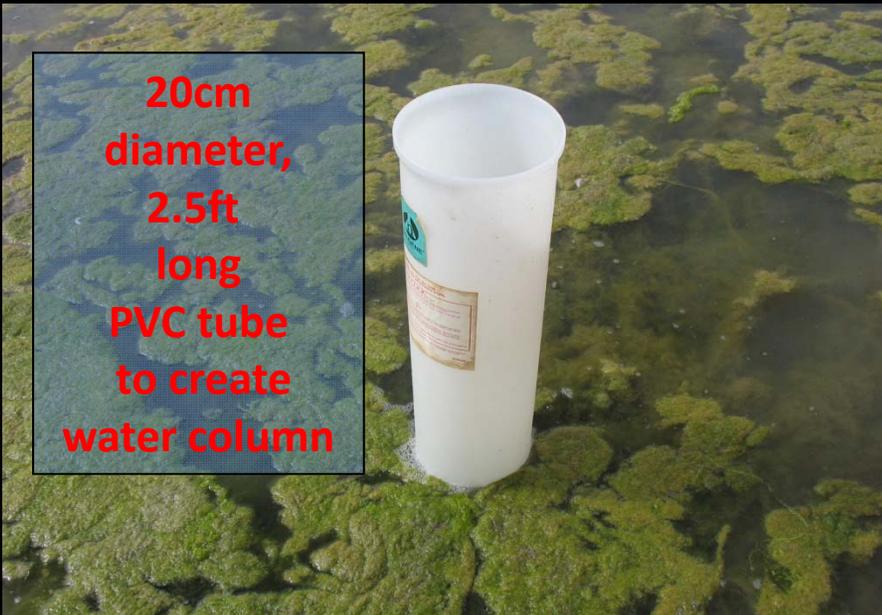
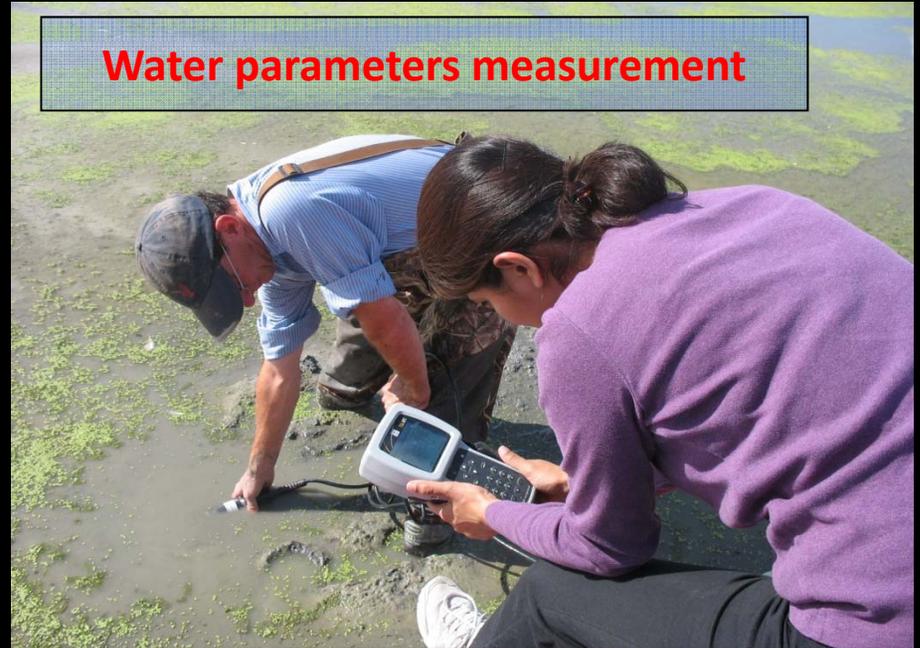
Microcystins, Nodularin, etc.

Anatoxins, Saxitoxin, etc.

Sample collection



Water parameters measurement



20cm diameter, 2.5ft long PVC tube to create water column

Scoop to collect water sample



Nodularin concentrations in monthly samples

2008: June, August, September, and November.

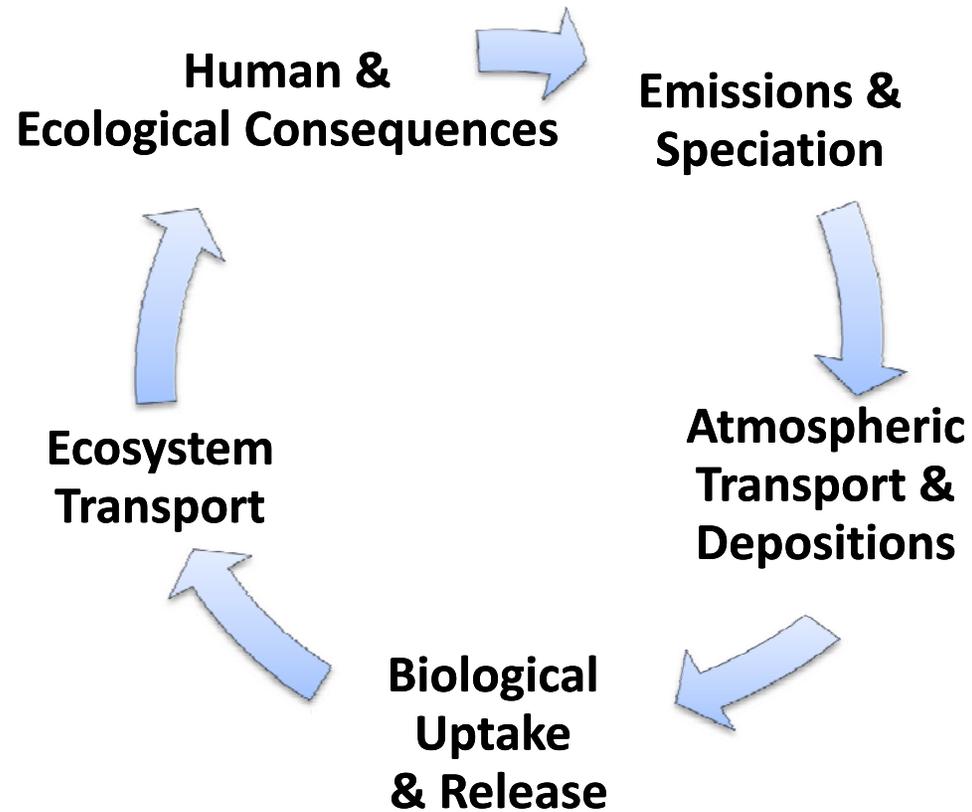
Table 1. Sampling sites gave positive Nodularin concentrations

Date	Site	Nodularin concentration ($\mu\text{g/L}$)
Aug.,08	4	0.384
	6	0.207
Sept.,08	4	0.331
	5	0.25

Conclusion: Nodularin was either completely absent or present in very low concentrations in the Farmington Bay well below $1\mu\text{g/L}$

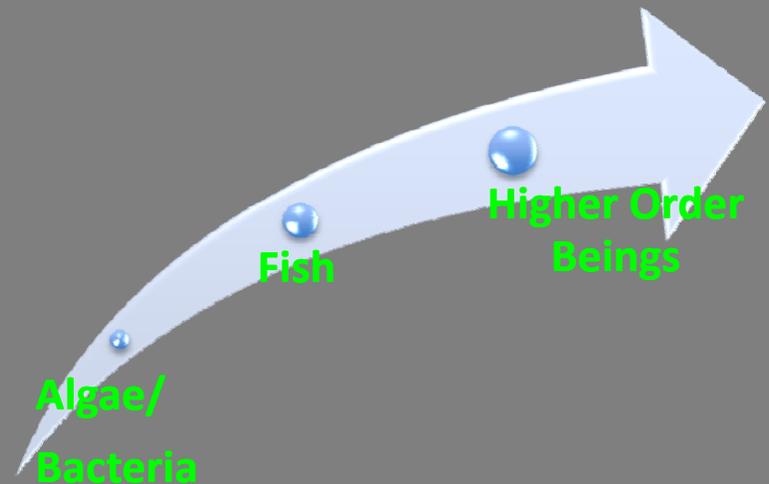
Mercury and its methylation

The Mercury Cycle



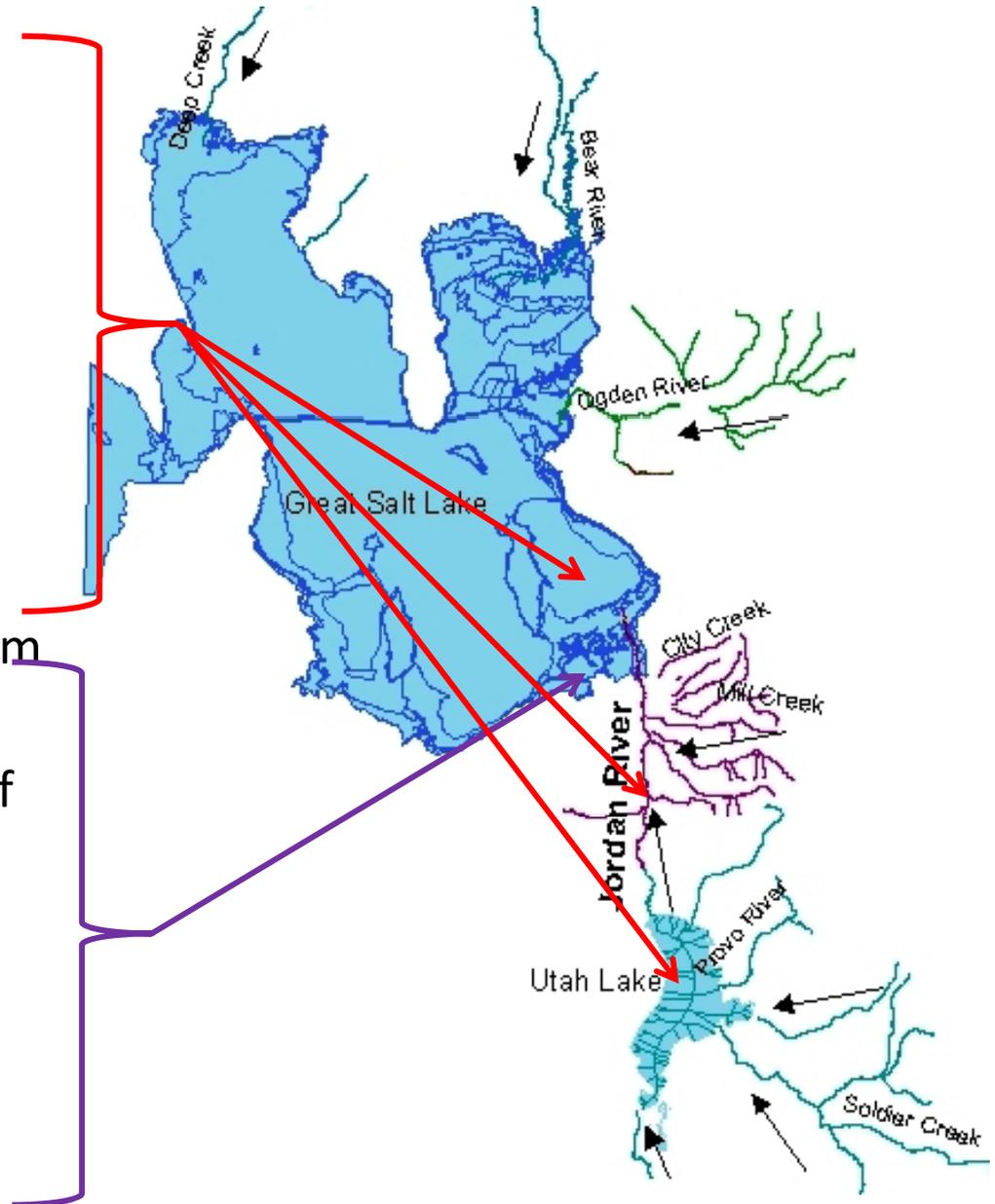
Concerns & Consequences

- Persistent Bioaccumulative Toxin (PBT)
- Ecological effects
 - Bioaccumulation & magnification
- Human health concerns
 - Potent neurotoxin
 - RfD for MeHg: $0.1 \mu\text{g}/\text{kg}\text{-day}$
 - Acute LD_{50} (Body weight of 70-kg human)
 - Inorganic: 14-57 mg/kg
 - Organic: 20-60 mg/kg (MeHg)



Research Objectives

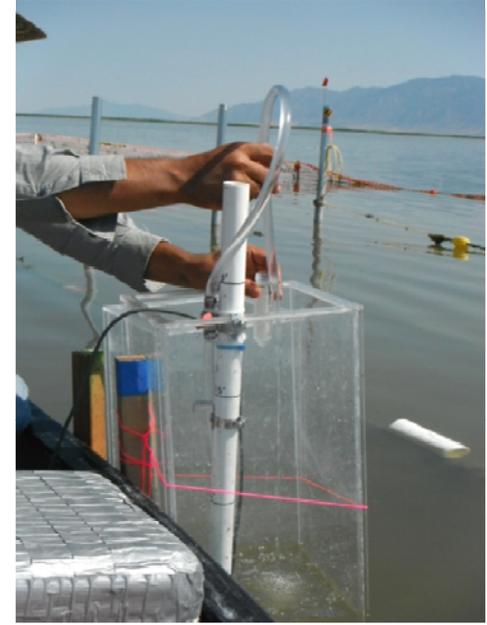
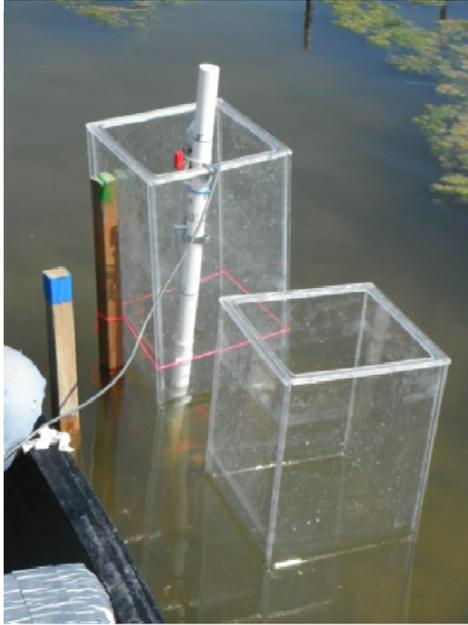
- Synoptic sampling & analysis of mercury, in water column & sediments from Farmington Bay (FB), Utah Lake and Jordan River
- Analyze the fate of mercury entering municipal waste water treatment plants (WWTP), emptying into Jordan River
- To evaluate the rate of mercury methylation in the sediments from the Turpin unit of FB
- And, to investigate the ecology of sulfate reducers possibly participating in mercury methylation of the sediments



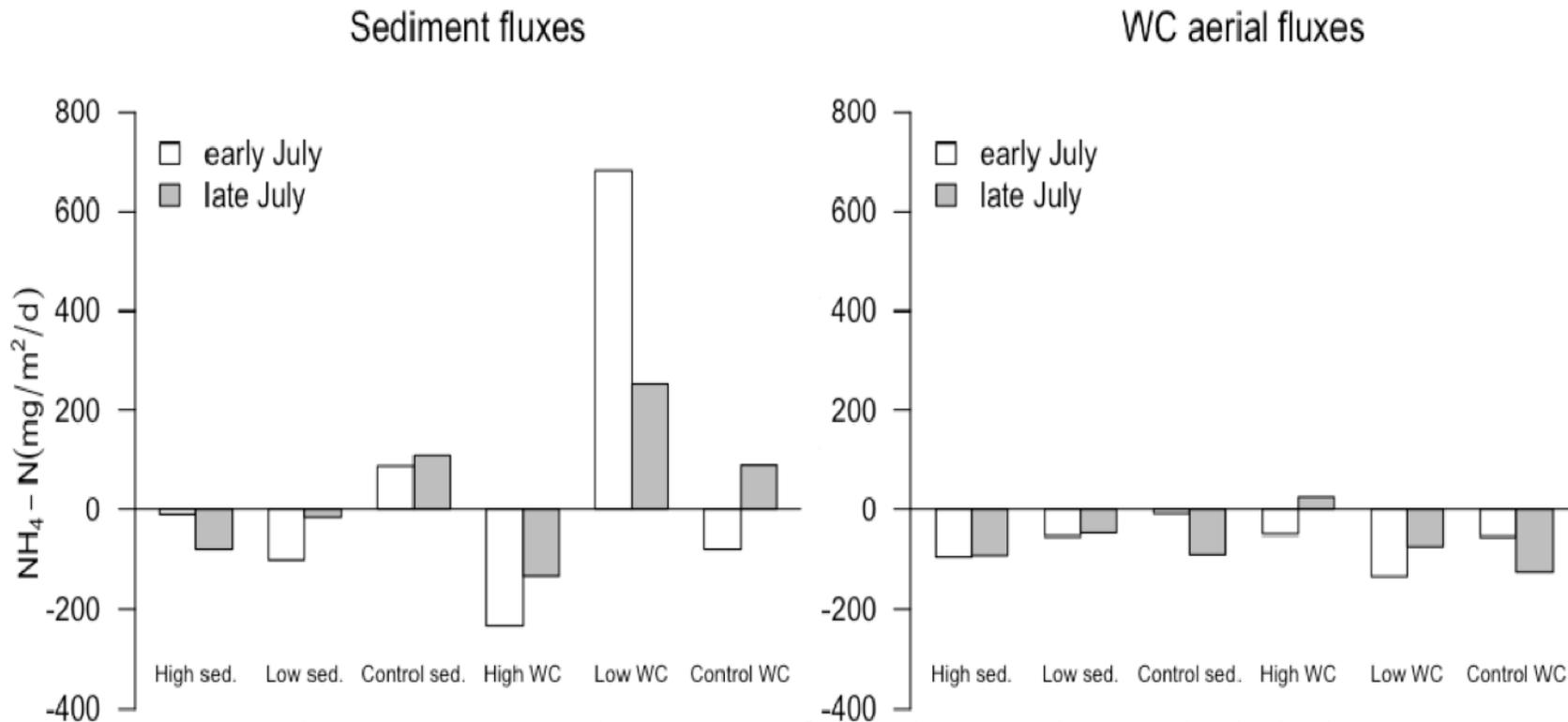
Summary of Analysis Results

Site	Matrix		Total Mercury	Methyl Mercury
FB	Water Column(ng/L)	Range	6.92-25.32	0.602-3.12
		Average	19.05	1.49
FB	Sediments (µg/Kg)	Range	34.01-83.1	0.06-1.51
		Average	60.47	0.35
Utah Lake	Water Column(ng/L)	Range	1.23-4.26	0.0055-0.7813
		Average	2.74	0.0536
Utah Lake	Sediments(µg/Kg)	Range	18-49	0.012-0.34
		Average	27.13	0.0717
Jordan River	Water Column(ng/L)	Range	19.17-26.33	0.10-0.0712
	Upper	Average	19.95	0.18
	Lower	Average	26.9	0.64
Jordan River	Sediments (µg/Kg)	Range	18-110	0.021-0.170
	Upper	Average	18.50	0.021
	Lower	Average	79.05	0.147

Willard Bay Nutrient Fluxes



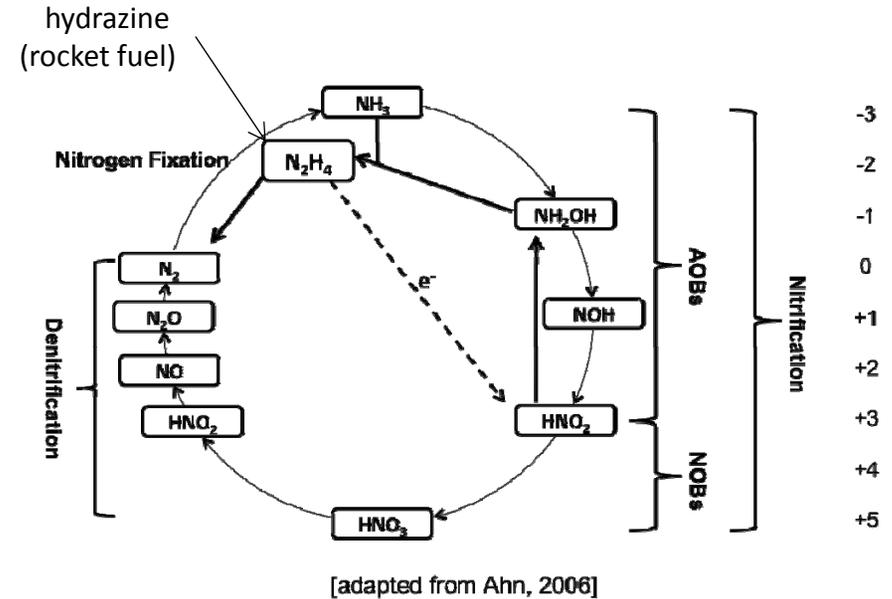
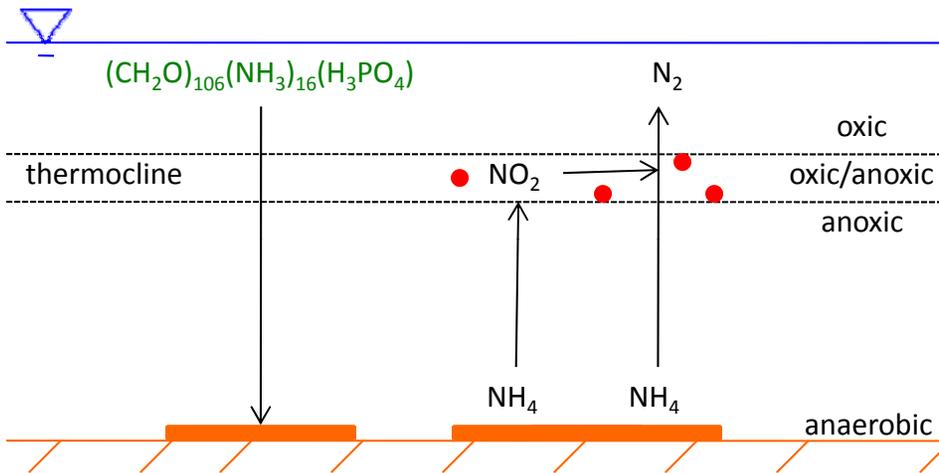
Sediment and WC fluxes- Ammonia



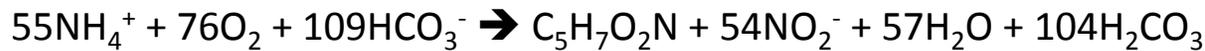
- Sediments removed ammonia from the WC during daylight hours
 - exceptions (Control Sediment & Low Water Column)
- WC removed ammonia during the daylight hours

Traditional Nitrogen Removal in Wetlands

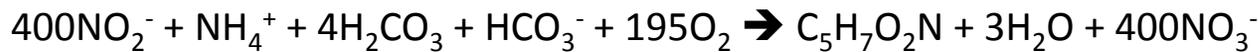
- Oceanographers noticed ammonium deficits in anoxic waters (Richards, 1965)
 - postulated using thermodynamics with molar ratios of $\text{NO}_2^-:\text{NH}_4^+$ of 1:1 and 1.67:1 (Broda, 1977)
 - 50-70% of N_2 gas produced in ocean
 - 30% of every breath we take



– Ammonia Oxidizing Bacteria (AOBs)



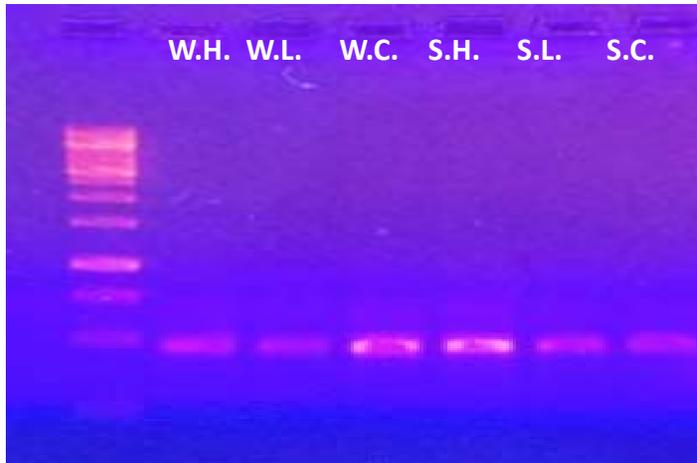
– Nitrite Oxidizing Bacteria (NOBs)



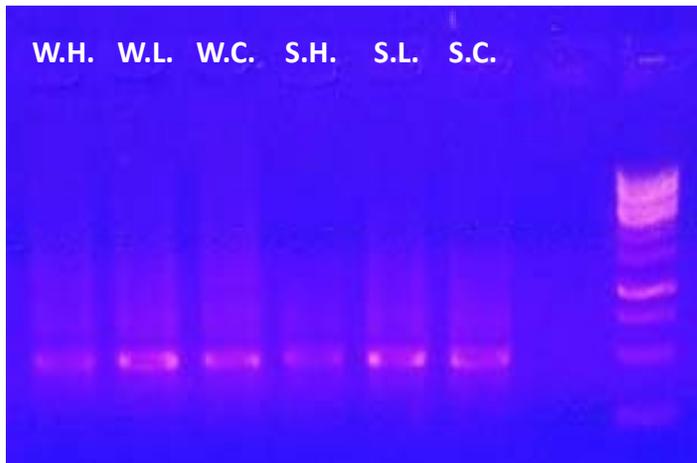
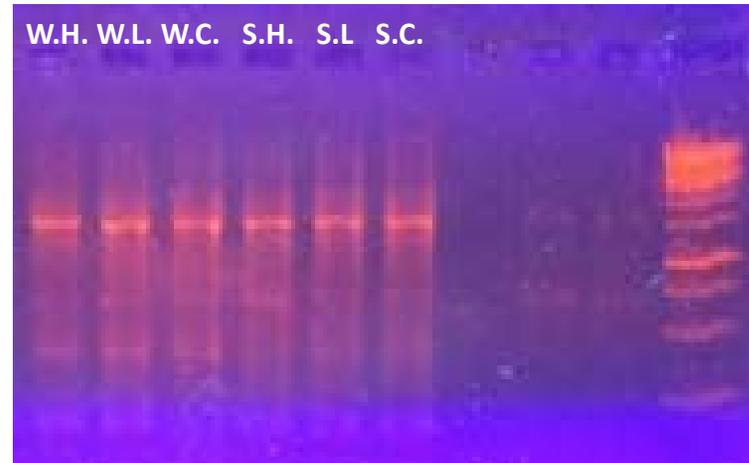
- $\text{NH}_4^+ + 1.32 \text{NO}_2^- + 0.066 \text{HCO}_3^- + 0.13 \text{H}^+ \rightarrow 1.02 \text{N}_2 + 0.26 \text{NO}_3^- + 0.066 \text{CH}_2\text{O}_{0.5}\text{N}_{0.15} + 2.03 \text{H}_2\text{O}$

Molecular Tests

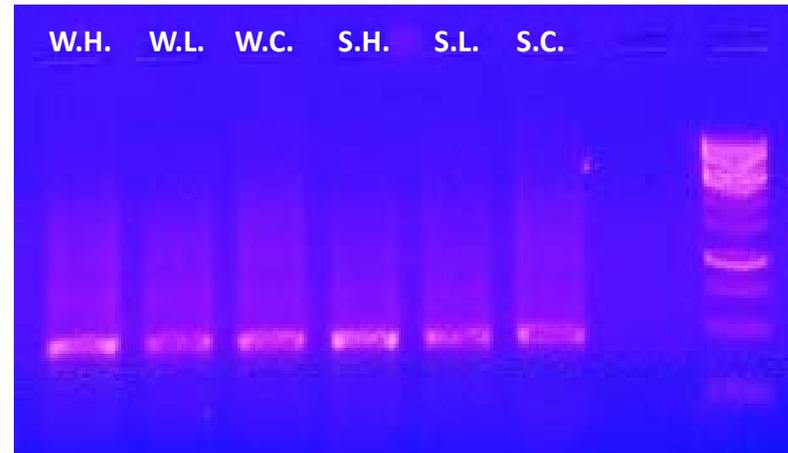
AOBs (amoA gene)
PCR (fragment size of **491 bp**)



All Anammox (1484 bp)



Brocadia and/or kueningenia
(452 bp)



Scalindua (452 bp)

QUESTIONS